

# Package ‘msdata’

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**Version** 0.32.0

**Title** Various Mass Spectrometry raw data example files

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**Depends** R (>= 2.10)

**Suggests** xcms, mzR, MSnbase

**ZipData** no

**Description** Ion Trap positive ionization mode data in mzData file format. Subset from 500-850 m/z and 1190-1310 seconds, incl. MS2 and MS3, intensity threshold 100.000. Extracts from FTICR Apex III, m/z 400-450. Subset of UPLC - Bruker micrOTOFq data, both mzData, mzML and mz5. LC-MSMS and MRM files from proteomics experiments. PSI mzIdentML example files for various search engines.

**biocViews** ExperimentData, MassSpectrometryData

**License** GPL (>= 2)

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msdata

*Sample FTICR, LC/MS and MS<sup>n</sup> data*

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## Description

x object containing a subset of LC/MS raw data from a Thermo Finnigan LCQ Deca XP The data is a subset from 500-850 m/z and 1190-1310 seconds, incl. MS2 and MS3, intensity threshold 100.000. It was collected in positive ionization mode.

xs object containing a subset of FTICR data from a Bruker APex III FTICR. The data is a subset from 400-450 m/z, collected in positive ionization mode.

## Usage

```
data(xs)
```

## Format

The format is:

xs

## Details

The corresponding raw mzdata files are located in the fticr and iontrap subdirectory of this package.

## See Also

[xcmsSet](#), [xcmsRaw](#)

## Examples

```
## The directory with the mzData LC/MS files
data(xs)
mzdatapath <- file.path(find.package("msdata"), "iontrap")
mzdatapath
files <- list.files(mzdatapath, recursive = TRUE, full.names = TRUE)
files

if (require(xcms)) {

  ## xcmsSet Summary
  show(xs)

  ## Access raw data file
  x <- xcmsRaw(files[1])
  x

}
```

proteomics

*Proteomics data in msdata*

## Description

This function returns proteomics mass spectrometry files. These files are all stored in the `proteomics` directory in the `msdata` package. Each file/data is described in more details below.

## Usage

```
proteomics(...)
```

## Arguments

... Additional arguments passed to `list.files`.

## Details

- `TMT_Erwinia_1uLSike_Top10HCD_isol2_45stepped_60min_01.mzML.gz`: A LC-MSMS data file containing TMT6 6-plex data. The data is described in more details in Gatto L. and Christoforou A. *Using R and Bioconductor for proteomics data analysis* (PMID [23692960](#)). This file only contains a subset of the full data (spectra 1002 to 1510) and was generated from the full data using `msconvert` (ProteoWizard release: 3.0.9283 (2016-1-11)) using following command

```
msconvert TMT_Erwinia_1uLSike_Top10HCD_isol2_45stepped_60min_01-20141210.mzML
--filter "index [1002,1510]" -o subset
```

The complete file is `TMT_Erwinia_1uLSike_Top10HCD_isol2_45stepped_60min_01-20141210.mzML.gz`, also available [here](#), and can also be downloaded from the ProteomeXchange PXD000001 project (see the `rpx` package).

An MS2 identification file, `ident/TMT_Erwinia_1uLSike_Top10HCD_isol2_45stepped_60min_01-20141210.mzML`, generated searching the raw data against the *Erwinia carotovora* database (see reference above) is also available through the `ident` function.

- `MS3TMT10_01022016_32917-33481.mzML.gz`: A subset of 565 spectra from a currently unpublished TMT 10-plex experiment run on an Thermo Orbitrap Lumos with synchronous precursor selection (SPS) MS3. Only the MS2 spectra were centroided during conversion using `msconvert` (ProteoWizard release: 3.0.9283 (2016-1-11)) using vendor libraries.
- `MS3TMT11.mzML`: A subset of 994 spectra from a currently unpublished MS3 SPS TMT 11-plex experiment converted to `mzML` using `msconvert`. The file contains 30, 482 and 482 MS1, MS2 and MS3 spectra, respectively. The MS1 spectra are in profile mode; other MS levels are centroided. See *Sensitive and Accurate Quantitation of Phosphopeptides Using TMT Isobaric Labeling Technique* for details about the acquisition method.

An feature data containing identification data is available with `data(fdms3tmt11)`, which can be used to directly update the feature data, as shown in the example below.

- `MRM-standmix-5.mzML.gz`: Sample from mouse brain acquired by HILIC ESI-QqQ/MS in Dynamic multiple reaction monitoring mode (MRM). HPLC system was a 1290 Infinity (Agilent Technologies) coupled to ion-Funnel Triple quadrupole 6490 mass spectrometer (Agilent Technologies). This file was contributed by Xavi Domingo-Almenara from the The Scripps Research Institute, San Diego, CA.

**Value**

A character with file names.

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**See Also**

For more access to mass spectrometry-based proteomics data, see the `rpx` and `ProteomicsAnnotationHubData` packages.

**Examples**

```
## raw data files
(f <- proteomics(full.names = TRUE))

library("mzR")
openMSfile(f[2])

library("MSnbase")
## The MS3 TMT11 raw data
(fms3 <- proteomics(full.names = TRUE, pattern = "MS3TMT11.mzML"))
ms3 <- readMSData(fms3, mode = "onDisk")
ms3

## Additional feature metadata
data(fdms3tmt11)
names(fdms3tmt11)

fData(ms3) <- fdms3tmt11
validObject(ms3)

## identification data file
ident(full.names = TRUE)

## quantiative data files
quant(full.names = TRUE)
```

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sciexdata

*AB Sciex LC-MS data files*

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**Description**

The mzML files in the `sciex` directory in the `msdata` package represent profile-mode LC-MS data of pooled human serum samples (the same pool being measured). The samples were analyzed by ultra high-performance liquid chromatography (UHPLC; Agilent 1290) coupled to a Q-TOF mass spectrometer (TripleTOF 5600+ AB Sciex). The chromatographic separation was based in hydrophilic interaction liquid chromatography (HILIC) and performed using an Waters Acquity BEH Amide, 100 x 2.1 mm column.

The mass spectrometer was operated in full scan mode in the mass range from 50 to 1000 m/z and with an accumulation time of 250 ms. The files represent a subset of spectra/scans from m/z 105 to 134 and from retention time 0 to 260 seconds. The files were generated in the same LC-MS run, but from different injections. Details on the individual files are provided below.

**Details**

- 20171016\_POOL\_POS\_1\_105-134.mzML profile-mode LC-MS data of pooled human serum samples. Injection index: 1.
- 20171016\_POOL\_POS\_3\_105-134.mzML profile-mode LC-MS data of pooled human serum samples. Injection index: 19.

**Author(s)**

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**Examples**

```
## List the files in the sciex folder  
dir(system.file("sciex", package = "msdata"))
```

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