Package 'Rbec'

October 10, 2021

Title Rbec: a tool for analysis of amplicon sequencing data from

Type Package

synthetic microbial communities
Version 1.0.0
Description Rbec is a adapted version of DADA2 for analyzing amplicon sequencing data from synthetic communities (SynComs), where the reference sequences for each strain exists. Rbec can not only accurately profile the microbial compositions in SynComs, but also predict the contaminants in SynCom samples.
License LGPL-3
Imports Rcpp (>= 1.0.6), dada2, ggplot2, readr, doParallel, foreach, grDevices, stats, utils
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Contam_detect

Reference-based error correction of amplicon sequencing data

Description

This function is designed for predicting the contaminated samples

Usage

```
Contam_detect(log_file, outdir, outlier_constant=1.5)
```

Arguments

```
log_file the file contains a list of log files of each sample outputted with Rbec function outdir output directory outlier_constant
```

the multiplier of variance to define the outlier

Details

Ruben Garrido-Oter's group, Plant-Microbe interaction, Max Planck Institute for Plant Breeding Research

Value

Returns a plot showing the distribution of percentage of corrected reads across the whole sample set and a summary file recording which samples might be contaminated

Author(s)

Pengfan Zhang

Examples

```
#log_file <- system.file("extdata", "rbec_test.list", package = "Rbec")
log_path <- list.files(paste(path.package("Rbec"),
   "extdata/contamination_test", sep="/"),
recursive=TRUE, full.names=TRUE)
log_file <- tempfile()
writeLines(log_path, log_file)
Contam_detect(log_file, tempdir())</pre>
```

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Rbec	Reference-based error correction of amplicon sequencing data

Description

This function corrects the amplicon sequencing data from synthetic communities where the reference sequences are known a priori

Usage

```
Rbec(fastq, reference, outdir, threads=1, sampling_size=5000, ascii=33, min_cont_abs=0.03)
```

Arguments

fastq the path of the fastq file containg merged amplicon sequencing reads (Ns are not

allowed in the reads)

reference the path of the unique reference sequences, each sequence must be in one line

(Ns are not allowed in the sequences)

outdir the output directory, which should be created by the user

threads the number of threads used, default 1

sampling_size the sampling size for calculating the error matrix, default 5000

ascii characters used to encode phred scores (33 or 64), default 33

min_cont_abs the relative abundance of unique tgas for detecting contamination sequences that

can't be corrected by any of the references

Details

Ruben Garrido-Oter's group, Plant-Microbe interaction, Max Planck Institute for Plant Breeding Research

Value

lambda_final.out the lambda value and pvalue of the Poisson distribution for each read error_matrix_final.out the error matrix in the final iteration strain_table.txt the strain composition of the sample contamination_seq.fna the potential sequences generated by contaminants rbec.log percentage of corrected reads, which can be used to predict contaminated samples

Author(s)

Pengfan Zhang

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Examples

```
fastq <- system.file("extdata", "test_raw_merged_reads.fastq.gz", package = "Rbec")
ref <- system.file("extdata", "test_ref.fasta", package = "Rbec")
Rbec(fastq=fastq, reference=ref, outdir=tempdir(), threads=1, sampling_size=500, ascii=33)</pre>
```

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