

# Package ‘GenomicDistributions’

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**Title** Produces Summaries and Plots of Features Distributed Across Genomes

**Description** If you have a set of genomic ranges, this package can help you with visualization and comparison. It produces several kinds of plots, for example: Chromosome distribution plots, which visualize how your regions are distributed over chromosomes; feature distance distribution plots, which visualizes how your regions are distributed relative to a feature of interest, like Transcription Start Sites (TSSs); genomic partition plots, which visualize how your regions overlap given genomic features such as promoters, introns, exons, or intergenic regions. It also makes it easy to compare one set of ranges to another.

**Depends** R ( $\geq 4.0$ ), IRanges, GenomicRanges

**Imports** data.table, ggplot2, reshape2, methods, utils, Biostrings,

**Suggests** AnnotationFilter, rtracklayer, testthat, knitr, BiocStyle, rmarkdown

**Enhances** BSgenome, extrafont, ensemblDb, GenomicFeatures

**LazyData** true

**VignetteBuilder** knitr

**License** BSD\_2\_clause + file LICENSE

**biocViews** Software, GenomeAnnotation, GenomeAssembly, DataRepresentation, Sequencing, Coverage, FunctionalGenomics, Visualization

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<code>.requireAndReturn</code>	<i>Checks to make sure a package object is installed, and if so, returns it. If the library is not installed, it issues a warning and returns NULL.</i>
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## Description

Checks to make sure a package object is installed, and if so, returns it. If the library is not installed, it issues a warning and returns NULL.

## Usage

```
.requireAndReturn(BSgenomeString)
```

## Arguments

`BSgenomeString` A BSgenome compatible genome string.

## Value

A BSgenome object if installed.

---

<code>.validateInputs</code>	<i>Checks class of the list of variables. To be used in functions</i>
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---

### Description

Checks class of the list of variables. To be used in functions

### Usage

```
.validateInputs(checkList)
```

### Arguments

<code>checkList</code>	list of object to check, e.g. <code>list(varname=c("data.frame", "numeric"))</code> . Multiuple strings in the vector are treated as OR.
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### Value

A warning if the wrong input class is provided.

### Examples

```
x <- function(var1) {
  cl = list(var1=c("numeric", "character"))
  .validateInputs(cl)
  return(var1^2)
}
```

---

<code>binBSGenome</code>	<i>Bins a BSgenome object.</i>
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### Description

Given a BSgenome object (to be loaded via `loadBSgenome`), and a number of bins, this will bin that genome. It is a simple wrapper of the `binChroms` function

### Usage

```
binBSGenome(genome, binCount)
```

### Arguments

<code>genome</code>	A UCSC-style string denoting reference assembly (e.g. 'hg38')
<code>binCount</code>	number of bins per chromosome

**Value**

A data.table object showing the region and bin IDs of the reference genome.

**Examples**

```
## Not run:
binCount = 1000
refGenomeBins = binBSGenome("hg19", binCount)

## End(Not run)
```

---

binChroms

*Naively splits a chromosome into bins*

---

**Description**

Given a list of chromosomes with corresponding sizes, this script will produce (roughly) evenly-sized bins across the chromosomes. It does not account for assembly gaps or the like.

**Usage**

```
binChroms(binCount, chromSizes)
```

**Arguments**

binCount	number of bins (total; *not* per chromosome)
chromSizes	a named list of size (length) for each chromosome.

**Value**

A data.table object assigning a bin ID to each chromosome region.

**Examples**

```
chromSizes = c(chr1=249250621, chr2=243199373, chr3=198022430)
cBins = binChroms(1000, chromSizes)
```

---

binRegion

---

*Divide regions into roughly equal bins*


---

### Description

Given a start coordinate, end coordinate, and number of bins to divide, this function will split the regions into that many bins. Bins will be only approximately the same size, due to rounding. (they should not be more than 1 different).

### Usage

```
binRegion(start, end, binSize = NULL, binCount = NULL, indicator = NULL)
```

### Arguments

start	The starting coordinate
end	The ending coordinate
binSize	The size of bin to divide the genome into. You must supply either binSize (priority) or binCount.
binCount	The number of bins to divide. If you do not supply binSize, you must supply binCount, which will be used to calculate the binSize.
indicator	A vector with identifiers to keep with your bins, in case you are doing this on a long table with multiple segments concatenated

### Details

Use case: take a set of regions, like CG islands, and bin them; now you can aggregate signal scores across the bins, giving you an aggregate signal in bins across many regions of the same type.

In theory, this just runs on 3 values, but you can run it inside a `data.table` `j` expression to divide a bunch of regions in the same way.

### Value

A `data.table`, expanded to `nrow` = number of bins, with these id columns: `id`: region ID `binID`: repeating ID (this is the value to aggregate across) `ubinID`: unique bin IDs

### Examples

```
Rbins = binRegion(1, 3000, 100, 1000)
```

---

BSdtToGRanges	<i>Converts a list of data.tables (From BSreadbeds) into GRanges.</i>
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---

**Description**

Converts a list of data.tables (From BSreadbeds) into GRanges.

**Usage**

```
BSdtToGRanges(dtList)
```

**Arguments**

dtList	A list of data.tables
--------	-----------------------

**Value**

A GRangesList object.

---

calcChromBins	<i>Calculates the distribution of a query set over the genome</i>
---------------	-------------------------------------------------------------------

---

**Description**

Returns a data.table showing counts of regions from the query that overlap with each bin. In other words, where on which chromosomes are the ranges distributed? You must provide binned regions. Only the midpoint of each query region is used to test for overlap with the bin regions.

**Usage**

```
calcChromBins(query, bins)
```

**Arguments**

query	A GenomicRanges or GenomicRangesList object with query regions
bins	Pre-computed bins (as a GRangesList object) to aggregate over; for example, these could be genome bins

**Value**

A data.table showing where on which chromosomes ranges are distributed.

**Examples**

```
vistaSftd = GenomicRanges::shift(vistaEnhancers, 100000)
vistaSftd2 = GenomicRanges::shift(vistaEnhancers, 200000)
calcChromBins(vistaEnhancers, GRangesList(vistaSftd, vistaSftd2))
```

---

calcChromBinsRef	<i>Returns the distribution of query over a reference assembly Given a query set of elements (a GRanges object) and a reference assembly (*e.g. 'hg38'), this will aggregate and count the distribution of the query elements across bins of the reference genome. This is a helper function to create features for common genomes. It is a wrapper of calcChromBins, which is more general.</i>
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---

### Description

Returns the distribution of query over a reference assembly Given a query set of elements (a GRanges object) and a reference assembly (\*e.g. 'hg38'), this will aggregate and count the distribution of the query elements across bins of the reference genome. This is a helper function to create features for common genomes. It is a wrapper of calcChromBins, which is more general.

### Usage

```
calcChromBinsRef(query, refAssembly, binCount = 10000)
```

### Arguments

query	A GenomicRanges or GenomicRangesList object with query regions
refAssembly	A character vector that will be used to grab chromosome sizes with getChromSizes
binCount	Number of bins to divide the chromosomes into

### Value

A data.table showing the distribution of regions across bins of the reference genome.

### Examples

```
ChromBins = calcChromBinsRef(vistaEnhancers, "hg19")
```

---

calcCumulativePartitions	<i>Calculates the cumulative distribution of overlaps between query and arbitrary genomic partitions</i>
--------------------------	----------------------------------------------------------------------------------------------------------

---

### Description

Takes a GRanges object, then assigns each element to a partition from the provided partitionList, and then tallies the number of regions assigned to each partition. A typical example of partitions is promoter, exon, intron, etc; this function will yield the number of each for a query GRanges object There will be a priority order to these, to account for regions that may overlap multiple genomic partitions.



**Usage**

```
calcCumulativePartitions(query, partitionList, remainder = "intergenic")
```

**Arguments**

query	GRanges or GRangesList with regions to classify.
partitionList	An ORDERED and NAMED list of genomic partitions GRanges. This list must be in priority order; the input will be assigned to the first partition it overlaps.
remainder	Which partition do you want to account for 'everything else'?

**Value**

A data.frame assigning each element of a GRanges object to a partition from a previously provided partitionList.

**Examples**

```
partitionList = genomePartitionList(geneModels_hg19$genesGR,
                                   geneModels_hg19$exonsGR,
                                   geneModels_hg19$threeUTRGR,
                                   geneModels_hg19$fiveUTRGR)
calcCumulativePartitions(vistaEnhancers, partitionList)
```

---

```
calcCumulativePartitionsRef
```

*Calculates the cumulative distribution of overlaps for a query set to a reference assembly*

---

**Description**

This function is a wrapper for calcCumulativePartitions that uses built-in partitions for a given reference genome assembly.

**Usage**

```
calcCumulativePartitionsRef(query, refAssembly)
```

**Arguments**

query	A GenomicRanges or GenomicRangesList object with query regions
refAssembly	A character vector specifying the reference genome assembly (*e.g.* 'hg19'). This will be used to grab chromosome sizes with getTSSs.

**Value**

A data.frame indicating the number of query region overlaps in several genomic partitions.

**Examples**

```
calcCumulativePartitionsRef(vistaEnhancers, "hg19")
```

---

```
calcExpectedPartitions
```

*Calculates the distribution of overlaps between query and arbitrary genomic partitions*

---

**Description**

Takes a GRanges object, then assigns each element to a partition from the provided partitionList, and then tallies the number of regions assigned to each partition. A typical example of partitions is promoter, exon, intron, etc; this function will yield the number of each for a query GRanges object. There will be a priority order to these, to account for regions that may overlap multiple genomic partitions.

**Usage**

```
calcExpectedPartitions(
  query,
  partitionList,
  genomeSize = NULL,
  remainder = "Intergenic"
)
```

**Arguments**

query	GRanges or GRangesList with regions to classify.
partitionList	An ORDERED and NAMED list of genomic partitions GRanges. This list must be in priority order; the input will be assigned to the first partition it overlaps.
genomeSize	The number of bases in the query genome. In other words, the sum of all chromosome sizes.
remainder	Which partition do you want to account for 'everything else'?

**Value**

A data.frame assigning each element of a GRanges object to a partition from a previously provided partitionList.

**Examples**

```
partitionList = genomePartitionList(geneModels_hg19$genesGR,
                                   geneModels_hg19$exonsGR,
                                   geneModels_hg19$threeUTRGR,
                                   geneModels_hg19$fiveUTRGR)

chromSizes = getChromSizes('hg19')
genomeSize = sum(chromSizes)
calcExpectedPartitions(vistaEnhancers, partitionList, genomeSize)
```

---

calcExpectedPartitionsRef

*Calculates the distribution of observed versus expected overlaps for a query set to a reference assembly*


---

### Description

This function is a wrapper for calcExpectedPartitions that uses built-in partitions for a given reference genome assembly.

### Usage

```
calcExpectedPartitionsRef(query, refAssembly)
```

### Arguments

query	A GenomicRanges or GenomicRangesList object with query regions
refAssembly	A character vector specifying the reference genome assembly (*e.g.* 'hg19'). This will be used to grab chromosome sizes with getTSSs.

### Value

A data.frame indicating the number of query region overlaps in several genomic partitions.

### Examples

```
calcExpectedPartitionsRef(vistaEnhancers, "hg19")
```

---

calcFeatureDist

*Find the distance to the nearest genomic feature*


---

### Description

For a given query set of genomic regions, and a given feature set of regions, this function will return the distance for each query region to its closest feature. It ignores strand and returns the distance as positive or negative, depending on whether the feature is upstream or downstream

### Usage

```
calcFeatureDist(query, features)
```

### Arguments

query	A GRanges or GRangesList object with query sets
features	A GRanges object with features to test distance to

**Details**

This function is similar to the bioconductor distanceToNearest function, but returns negative values for downstream distances instead of absolute values. This allows you to assess the relative location.

**Value**

A vector of genomic distances for each query region relative to its closest feature.

**Examples**

```
vistaSftd = GenomicRanges::shift(vistaEnhancers, 100000)
calcFeatureDist(vistaEnhancers, vistaSftd)
```

---

calcFeatureDistRefTSS *Calculates the distribution of distances from a query set to closest TSS*

---

**Description**

Given a query GRanges object and an assembly string, this function will grab the TSS list for the given reference assembly and then calculate the distance from each query feature to the closest TSS. It is a wrapper of calcFeatureDist that uses built-in TSS features for a reference assembly

**Usage**

```
calcFeatureDistRefTSS(query, refAssembly)
```

**Arguments**

query	A GenomicRanges or GenomicRangesList object with query regions
refAssembly	A character vector specifying the reference genome assembly (*e.g.* 'hg19'). This will be used to grab chromosome sizes with getTSSs.

**Value**

A vector of distances for each query region relative to TSSs.

**Examples**

```
calcFeatureDistRefTSS(vistaEnhancers, "hg19")
```

---

calcGCCContent	<i>Calculate GC content over genomic ranges</i>
----------------	-------------------------------------------------

---

**Description**

Given a reference genome as a BSgenome object and some ranges on that reference, this function will return a vector of the same length as the granges object, with percent of Cs and Gs.

**Usage**

```
calcGCCContent(query, ref)
```

**Arguments**

query	A GenomicRanges or GenomicRangesList object with query regions.
ref	Reference genome BSgenome object.

**Value**

A numeric vector of list of vectors with the GC percentage of the query regions.

**Examples**

```
## Not run:
bsg = loadBSgenome('hg19')
gcvec = calcGCCContent(vistaEnhancers, bsg)

## End(Not run)
```

---

calcGCCContentRef	<i>Calculate GC content over genomic ranges</i>
-------------------	-------------------------------------------------

---

**Description**

Given a reference genome as a BSgenome object and some ranges on that reference, this function will return a vector of the same length as the granges object, with percent of Cs and Gs.

**Usage**

```
calcGCCContentRef(query, refAssembly)
```

**Arguments**

query	A GenomicRanges or GenomicRangesList object with query regions
refAssembly	A character vector specifying the reference genome assembly (*e.g.* 'hg19'). This will be used to grab chromosome sizes with getTSSs.

**Value**

A numeric vector or list of vectors with the GC percentage of the query regions.

**Examples**

```
## Not run:  
refAssembly = 'hg19'  
GCcontent = calcGCContentRef(vistaEnhancers, refAssembly)  
  
## End(Not run)
```

---

calcNeighborDist	<i>Group regions from the same chromosome together and calculate the distances between neighboring regions. Distances are then lumped into a numeric vector.</i>
------------------	------------------------------------------------------------------------------------------------------------------------------------------------------------------

---

**Description**

Group regions from the same chromosome together and calculate the distances between neighboring regions. Distances are then lumped into a numeric vector.

**Usage**

```
calcNeighborDist(query)
```

**Arguments**

query            A GRanges or GRangesList object.

**Value**

A numeric vector or list with different vectors containing the distances within neighboring regions.

**Examples**

```
dist = calcNeighborDist(vistaEnhancers)
```

---

calcOpenSignal	<i>The function calcOpenSignal takes the input BED file(s) in form of GRanges or GRangesList object, overlaps it with all defined open chromatin regions across cell types and returns a matrix, where each row is the input genomic region (if overlap was found), each column is a cell type, and the value is a normalized ATAC-seq signal.</i>
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---

### Description

The function calcOpenSignal takes the input BED file(s) in form of GRanges or GRangesList object, overlaps it with all defined open chromatin regions across cell types and returns a matrix, where each row is the input genomic region (if overlap was found), each column is a cell type, and the value is a normalized ATAC-seq signal.

### Usage

```
calcOpenSignal(query, cellMatrix)
```

### Arguments

query	Genomic regions to be analyzed. Can be GRanges or GRangesList object.
cellMatrix	Matrix with open chromatin signal values, rows are genomic regions, columns are cell types. First column contains information about the genomic region in following form: chr_start_end. Can be either data.frame or data.table object.

### Value

A list with named components: signalMatrix - data.table with cell specific open chromatin signal values for query regions matrixStats - data.frame containing boxplot stats for individual cell type

### Examples

```
openRegionSummary = calcOpenSignal(vistaEnhancers, exampleOpenSignalMatrix_hg19)
```

---

calcPartitions	<i>Calculates the distribution of overlaps between query and arbitrary genomic partitions</i>
----------------	-----------------------------------------------------------------------------------------------

---

### Description

Takes a GRanges object, then assigns each element to a partition from the provided partitionList, and then tallies the number of regions assigned to each partition. A typical example of partitions is promoter, exon, intron, etc; this function will yield the number of each for a query GRanges object. There will be a priority order to these, to account for regions that may overlap multiple genomic partitions.

**Usage**

```
calcPartitions(query, partitionList, remainder = "intergenic")
```

**Arguments**

query	GRanges or GRangesList with regions to classify
partitionList	an ORDERED and NAMED list of genomic partitions GRanges. This list must be in priority order; the input will be assigned to the first partition it overlaps
remainder	A character vector to assign any query regions that do not overlap with anything in the partitionList. Defaults to "intergenic"

**Value**

A data.frame assigning each element of a GRanges object to a partition from a previously provided partitionList.

**Examples**

```
partitionList = genomePartitionList(geneModels_hg19$genesGR,
                                   geneModels_hg19$exonsGR,
                                   geneModels_hg19$threeUTRGR,
                                   geneModels_hg19$fiveUTRGR)
calcPartitions(vistaEnhancers, partitionList)
```

---

calcPartitionsRef	<i>Calculates the distribution of overlaps for a query set to a reference assembly</i>
-------------------	----------------------------------------------------------------------------------------

---

**Description**

This function is a wrapper for calcPartitions and calcPartitionPercents that uses built-in partitions for a given reference genome assembly.

**Usage**

```
calcPartitionsRef(query, refAssembly)
```

**Arguments**

query	A GenomicRanges or GenomicRangesList object with query regions
refAssembly	A character vector specifying the reference genome assembly (*e.g.* 'hg19'). This will be used to grab chromosome sizes with getTSSs.

**Value**

A data.frame indicating the number of query region overlaps in several genomic partitions.



Examples

```
calcPartitionsRef(vistaEnhancers, "hg19")
```

---

calcWidth	<i>Calculate the widths of regions</i>
-----------	----------------------------------------

---

Description

The length of a genomic region (the distance between the start and end) is called the width When given a query set of genomic regions, this function returns the width

Usage

```
calcWidth(query)
```

Arguments

query                    A GRanges or GRangesList object with query sets

Value

A vector of the widths (end-start coordinates) of GRanges objects.

Examples

```
TSSdist = calcFeatureDistRefTSS(vistaEnhancers, "hg19")
plotFeatureDist(TSSdist, featureName="TSS")
```

---

cellTypeMetadata	<i>Table the maps cell types to tissues and groups</i>
------------------	--------------------------------------------------------

---

Description

Table the maps cell types to tissues and groups

Usage

```
data(cellTypeMetadata)
```

Format

data.table with 3 columns (cellType, tissue and group) and 74 rows (one per cellType)

Source

self-curated dataset

---

chromSizes_hg19	<i>hg19 chromosome sizes</i>
-----------------	------------------------------

---

### Description

A dataset containing chromosome sizes for Homo Sapiens hg38 genome assembly

### Usage

```
data(chromSizes_hg19)
```

### Format

A named vectors of lengths with one item per chromosome

### Source

BSgenome.Hsapiens.UCSC.hg19 package

---

dtToGr	<i>Converts a data.table (DT) object to a GenomicRanges (GR) object. Tries to be intelligent, guessing chr and start, but you have to supply end or other columns if you want them to be carried into the GR.</i>
--------	-------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------

---

### Description

Converts a data.table (DT) object to a GenomicRanges (GR) object. Tries to be intelligent, guessing chr and start, but you have to supply end or other columns if you want them to be carried into the GR.

### Usage

```
dtToGr(
  DT,
  chr = "chr",
  start = "start",
  end = NA,
  strand = NA,
  name = NA,
  splitFactor = NA,
  metaCols = NA
)
```

**Arguments**

DT	A data.table representing genomic regions.
chr	A string representing the chromosome column.
start	A string representing the name of the start column.
end	A string representing the name of the end column.
strand	A string representing the name of the strand column.
name	A string representing the name of the name column.
splitFactor	A string representing the name of the column to use to split the data.table into multiple data.tables.
metaCols	A string representing the name of the metadata column(s) to include in the returned GRanges object.

**Value**

A GRanges object.

**Examples**

```
start1 = c(seq(from=1, to = 2001, by = 1000), 800)
chrString1 = c(rep("chr1", 3), "chr2")
dt = data.table::data.table(chr=chrString1,
                           start=start1,
                           end=start1 + 250)

newGR = dtToGr(dt)
```

---

dtToGrInternal	<i>Two utility functions for converting data.tables into GRanges objects</i>
----------------	------------------------------------------------------------------------------

---

**Description**

Two utility functions for converting data.tables into GRanges objects

**Usage**

```
dtToGrInternal(DT, chr, start, end = NA, strand = NA, name = NA, metaCols = NA)
```

**Arguments**

DT	A data.table representing genomic regions.
chr	A string representing the chromosome column.
start	A string representing the name of the start column.
end	A string representing the name of the end column.
strand	A string representing the name of the strand column.
name	A string representing the name of the name column.
metaCols	A string representing the name of the metadata column(s) to include in the returned GRanges object.

**Value**

A GRanges object.

---

exampleOpenSignalMatrix\_hg19

*A dataset containing a subset of open chromatin regions across all cell types defined by ENCODE for Homo Sapiens hg19*

---

**Description**

Preparation steps:

1. made a universe of regions by merging regions across cell types defined as opened in ENCODE
2. took bigwig files from ENCODE for individual cell types, merged replicates, filtered out blacklisted sites
3. evaluated the signal above regions defined by previous step
4. performed quantile normalization
5. subsetting it

**Usage**

```
data(exampleOpenSignalMatrix_hg19)
```

**Format**

data.frame, rows represent whole selection of open chromatin regions across all cell types defined by ENCODE, columns are individual cell types and values are normalized open chromatin signal values.

**Source**

[http://big.databio.org/open\\_chromatin\\_matrix/openSignalMatrix\\_hg19\\_quantileNormalized\\_round4.txt.gz](http://big.databio.org/open_chromatin_matrix/openSignalMatrix_hg19_quantileNormalized_round4.txt.gz)

---

geneModels_hg19	<i>hg38 gene models</i>
-----------------	-------------------------

---

**Description**

A dataset containing gene models for Homo Sapiens hg38 genome assembly.

**Usage**

```
data(geneModels_hg19)
```

**Format**

A list of two GRanges objects, with genes and exons locations

**Source**

EnsDb.Hsapiens.v75 package

---

genomePartitionList	<i>Create a basic genome partition list of genes, exons, introns, UTRs, and intergenic</i>
---------------------	--------------------------------------------------------------------------------------------

---

**Description**

Given GRanges for genes, and a GRanges for exons, returns a list of GRanges corresponding to various breakdown of the genome, based on the given annotations; it gives you proximal and core promoters, exons, and introns.

**Usage**

```
genomePartitionList(genesGR, exonsGR, threeUTRGR = NULL, fiveUTRGR = NULL)
```

**Arguments**

genesGR	a GRanges object of gene coordinates
exonsGR	a GRanges object of exons coordinates
threeUTRGR	a GRanges object of 3' UTRs
fiveUTRGR	a GRanges object of 5' UTRs

**Details**

To be used as a partitionList for calcPartitions.

**Value**

A list of GRanges objects, each corresponding to a partition of the genome. Partitions include proximal and core promoters, exons and introns.

**Examples**

```
partitionList = genomePartitionList(geneModels_hg19$genesGR,
                                     geneModels_hg19$exonsGR,
                                     geneModels_hg19$threeUTRGR,
                                     geneModels_hg19$fiveUTRGR)
```

---

GenomicDistributions    *Produces summaries and plots of features distributed across genomes*

---

**Description**

If you have a set of genomic ranges, the GenomicDistributions R package can help you with some simple visualizations. Currently, it can produce two kinds of plots: First, the chromosome distribution plot, which visualizes how your regions are distributed over chromosomes; and second, the feature distribution plot, which visualizes how your regions are distributed relative to a feature of interest, like Transcription Start Sites (TSSs).

**Author(s)**

Nathan C. Sheffield

**References**

<http://github.com/databio/GenomicDistributions>

---

getChromSizes    *Returns built-in chrom sizes for a given reference assembly*

---

**Description**

Returns built-in chrom sizes for a given reference assembly

**Usage**

```
getChromSizes(refAssembly)
```

**Arguments**

refAssembly    A string identifier for the reference assembly

**Value**

A vector with the chromosome sizes corresponding to a specific genome assembly.

**Examples**

```
getChromSizes("hg19")
```

---

getGeneModels

*Returns built-in gene models for a given reference assembly*

---

**Description**

Some functions require gene models, which can be obtained from any source. This function allows you to retrieve a few common built-in ones.

**Usage**

```
getGeneModels(refAssembly)
```

**Arguments**

refAssembly     A string identifier for the reference assembly

**Value**

A list containing the gene models corresponding to a specific reference assembly.

**Examples**

```
getGeneModels("hg19")
```

---

getReferenceData

*Get reference data for a specified assembly*

---

**Description**

This is a generic getter function that will return a data object requested, if it is included in the built-in data with the GenomicDistributions package or GenomicDistributionsData package (if installed). Data objects can be requested for different reference assemblies and data types (specified by a tagline, which is a unique string identifying the data type).

**Usage**

```
getReferenceData(refAssembly, tagline)
```

**Arguments**

refAssembly	Reference assembly string (e.g. 'hg38')
tagline	The string that was used to identify data of a given type in the data building step. It's used for the filename so we know what to load, and is what makes this function generic (so it can load different data types).

**Value**

A requested and included package data object.

---

grToDt	<i>Convert a GenomicRanges into a data.table.</i>
--------	---------------------------------------------------

---

**Description**

Convert a GenomicRanges into a data.table.

**Usage**

```
grToDt(GR)
```

**Arguments**

GR	A Granges object
----	------------------

**Value**

A data.table object.

---

labelCuts	<i>Creates labels based on a discretization definition.</i>
-----------	-------------------------------------------------------------

---

**Description**

If you are building a histogram of binned values, you want to have labels for your bins that correspond to the ranges you used to bin. This function takes the breakpoints that define your bins and produces nice-looking labels for your histogram plot.

**Usage**

```
labelCuts(
  breakpoints,
  round_digits = 1,
  signif_digits = 3,
  collapse = "-",
  infBins = FALSE
)
```



**Arguments**

breakPoints	The exact values you want as boundaries for your bins
round_digits	Number of digits to cut round labels to.
signif_digits	Number of significant digits to specify.
collapse	Character to separate the labels
infBins	use >/< as labels on the edge bins

**Details**

labelCuts will take a cut group, (e.g., a quantile division of some signal), and give you clean labels (similar to the cut method).

**Value**

A vector of histogram axis labels.

---

loadBSgenome	<i>Loads BSgenome objects from UCSC-style character vectors.</i>
--------------	------------------------------------------------------------------

---

**Description**

This function will let you use a simple character vector (e.g. 'hg19') to load and then return BSgenome objects. This lets you avoid having to use the more complex annotation for a complete BSgenome object (e.g. BSgenome.Hsapiens.UCSC.hg38.masked)

**Usage**

```
loadBSgenome(genomeBuild, masked = TRUE)
```

**Arguments**

genomeBuild	One of 'hg19', 'hg38', 'mm10', 'mm9', or 'grch38'
masked	Should we used the masked version? Default:TRUE

**Value**

A BSgenome object corresponding to the provided genome build.

**Examples**

```
## Not run:
bsg = loadBSgenome('hg19')

## End(Not run)
```

---

loadEnsDb	<i>Load selected EnsDb library</i>
-----------	------------------------------------

---

**Description**

Load selected EnsDb library

**Usage**

```
loadEnsDb(genomeBuild)
```

**Arguments**

genomeBuild      string, genome identifier

**Value**

loaded library

**Examples**

```
## Not run:
loadEnsDb("hg19")

## End(Not run)
```

---

neighborDt	<i>Internal helper function to calculate distance between neighboring regions.</i>
------------	------------------------------------------------------------------------------------

---

**Description**

Internal helper function to calculate distance between neighboring regions.

**Usage**

```
neighborDt(queryDt)
```

**Arguments**

queryDt            A data table with regions grouped according to chromosome.

**Value**

A numeric vector with the distances in bp

---

nlist	<i>Nathan's magical named list function. This function is a drop-in replacement for the base list() function, which automatically names your list according to the names of the variables used to construct it. It seamlessly handles lists with some names and others absent, not overwriting specified names while naming any unnamed parameters. Took me awhile to figure this out.</i>
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---

**Description**

Nathan's magical named list function. This function is a drop-in replacement for the base list() function, which automatically names your list according to the names of the variables used to construct it. It seamlessly handles lists with some names and others absent, not overwriting specified names while naming any unnamed parameters. Took me awhile to figure this out.

**Usage**

```
nlist(...)
```

**Arguments**

```
...           arguments passed to list()
```

**Value**

A named list object.

**Examples**

```
x=5
y=10
nlist(x,y) # returns list(x=5, y=10)
list(x,y) # returns unnamed list(5, 10)
```

---

plotChromBins	<i>Plot distribution over chromosomes</i>
---------------	-------------------------------------------

---

**Description**

Plots result from genomicDistribution calculation

**Usage**

```
plotChromBins(
  genomeAggregate,
  binCount = 10000,
  plotTitle = "Distribution over chromosomes"
)
```

**Arguments**

genomeAggregate	The output from the genomicDistribution function
binCount	Number of bins (should match the call to genomicDistribution)
plotTitle	Title for plot.

**Value**

A ggplot object showing the distribution of the query regions over bins of the reference genome.

**Examples**

```
agg = data.frame("regionID"=1:5, "chr"=rep(c("chr1"), 5),
                 "withinGroupID"=1:5, "N"=c(1,3,5,7,9))
ChromBins = plotChromBins(agg)
```

---

plotCumulativePartitions

*Plot the cumulative distribution of regions in features*

---

**Description**

This function plots the cumulative distribution of regions across a feature set.

**Usage**

```
plotCumulativePartitions(assignedPartitions, feature_names = NULL)
```

**Arguments**

assignedPartitions	Results from calcCumulativePartitions
feature_names	An optional character vector of feature names, in the same order as the GenomicRanges or GenomicRangesList object.

**Value**

A ggplot object of the cumulative distribution of regions in features.

**Examples**

```
p = calcCumulativePartitionsRef(vistaEnhancers, "hg19")
cumuPlot = plotCumulativePartitions(p)
```

---

`plotExpectedPartitions`

*Produces a barplot showing how query regions of interest are distributed relative to the expected distribution across a given partition list*

---

### Description

Produces a barplot showing how query regions of interest are distributed relative to the expected distribution across a given partition list

### Usage

```
plotExpectedPartitions(expectedPartitions, feature_names = NULL)
```

### Arguments

`expectedPartitions`

A data.frame holding the frequency of assignment to each of the partitions, the expected number of each partition, and the log10 of the observed over expected. Produced by `calcExpectedPartitions`.

`feature_names`

Character vector with labels for the partitions (optional). By default it will use the names from the first argument.

### Value

A ggplot object using a barplot to show the distribution of the query regions across a given partition list.

### Examples

```
p = calcExpectedPartitionsRef(vistaEnhancers, "hg19")
expectedPlot = plotExpectedPartitions(p)
```

---

`plotFeatureDist`

*Plots a histogram of distances to genomic features*

---

### Description

Given the results from `featureDistribution`, plots a histogram of distances surrounding the features of interest

**Usage**

```
plotFeatureDist(
  dists,
  bgdists = NULL,
  featureName = "features",
  numbers = FALSE,
  nbins = 50,
  size = 1e+05,
  infBins = FALSE,
  tile = FALSE
)
```

**Arguments**

dists	Results from featureDistribution
bgdists	Background distances. If provided, will plot a background distribution of expected distances
featureName	Character vector for plot labels (optional).
numbers	a logical indicating whether the raw numbers should be displayed, rather than percentages (optional).
nbins	Number of bins on each side of the center point.
size	Number of bases to include in plot on each side of the center point.
infBins	Include catch-all bins on the sides?
tile	Turn on a tile mode, which plots a tiled figure instead of a histogram.

**Value**

A ggplot2 plot object

**Examples**

```
TSSdist = calcFeatureDistRefTSS(vistaEnhancers, "hg19")
f = plotFeatureDist(TSSdist, featureName="TSS")
```

---

plotGCCContent	<i>Plots a density distribution of GC vectors Give results from the calcGCCContent function, this will produce a density plot</i>
----------------	-----------------------------------------------------------------------------------------------------------------------------------

---

**Description**

Plots a density distribution of GC vectors Give results from the calcGCCContent function, this will produce a density plot

**Usage**

```
plotGCCContent(gcvecs)
```

**Arguments**

gcvectors      A numeric vector or list of numeric vectors of GC contents.

**Value**

A ggplot object plotting distribution of GC content in query regions.

**Examples**

```
numVector = rnorm(400, mean=0.5, sd=0.1)
GCplot = plotGCCContent(numVector)
vecs = list(example1 = rnorm(400, mean=0.5, sd=0.1),
            example2 = rnorm(600, mean=0.5, sd=0.1))
GCplot = plotGCCContent(numVector)
```

---

plotNeighborDist	<i>Plot the distances between neighboring regions. The distance in the x axis is log10 transformed for ease of comparison between different regionsets and to account for outliers.</i>
------------------	-----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------

---

**Description**

Plot the distances between neighboring regions. The distance in the x axis is log10 transformed for ease of comparison between different regionsets and to account for outliers.

**Usage**

```
plotNeighborDist(dcvec)
```

**Arguments**

dcvec      A numeric vector or list with vectors containing distances between neighbor regions. Produced by calcNeighborDist

**Value**

A ggplot density object showing the distribution of log10 transformed distances.

**Examples**

```
numVector = rnorm(400, mean=5, sd=0.1)
d = plotNeighborDist(numVector)
```

---

plotOpenSignal	<i>The function plotOpenSignal visualizes the signalMatrix obtained from calcOpenSignal.</i>
----------------	----------------------------------------------------------------------------------------------

---

## Description

The function plotOpenSignal visualizes the signalMatrix obtained from calcOpenSignal.

## Usage

```
plotOpenSignal(
  openRegionSummary,
  plotType = "barPlot",
  cellGroup = NA,
  cellTypeMetadata = NA,
  colorScheme = c("#E31A1C", "#666666", "#B3DE69", "#A65628", "#33A02C", "#E6AB02",
    "#F0027F", "#FDC086", "#FFFF99", "#B3E2CD", "#B3CDE3", "#66A61E", "#F4CAE4",
    "#80B1D3", "#FFED6F", "#B15928", "#999999")
)
```

## Arguments

openRegionSummary	Output list from calcOpenSignal function.
plotType	Options are: jitter - jitter plot with box plot on top boxPlot - box plot without individual points and outliers barPlot (default) - bar height represents the median signal value for a given cell type.
cellGroup	- This option allows to select a tissue type to be plotted, if NA (default) all available tissue types are plotted, available options: "blood", "bone", "CNS", "embryonic", "eye", "foreskin", "gastrointestinal", "heart", "liver", "lymphatic", "mammaryGland", "mouth", "respiratorySystem", "skeletalMuscle", "skin", "urinarySystem", "vasculature", can be passed as a character string or vector of strings.
cellTypeMetadata	Metadata for cell type - tissue association. This option is for users, who provide their own open region signal matrix. The cellTypeMetadata matrix must contain two columns called cellType and tissueType. cellType column contains the cell type names in the provided signalMatrix column names. The tissueType columns provides an information, which tissue the cell type comes from.
colorScheme	Provide color values for each tissueType if you want to change the default colors.

## Value

A ggplot object.



**Examples**

```
## Not run:
openRegionSummary = calcOpenSignal(vistaEnhancers, exampleOpenSignalMatrix_hg19)
plotSignal = plotOpenSignal(openRegionSummary)
plotSignal = plotOpenSignal(openRegionSummary, plotType = "jitter",
cellGroup = "blood")

## End(Not run)
```

---

plotPartitions	<i>Produces a barplot showing how query regions of interest are distributed across a given partition list</i>
----------------	---------------------------------------------------------------------------------------------------------------

---

**Description**

This function can be used to test a GRanges object against any arbitrary list of genome partitions. The partition list is a priority-ordered list of GRanges objects. Each region in the query will be assigned to a given partition that it overlaps with the highest priority.

**Usage**

```
plotPartitions(assignedPartitions, numbers = FALSE)
```

**Arguments**

assignedPartitions	A table holding the frequency of assignment to each of the partitions. Produced by calcPartitions
numbers	logical indicating whether raw overlaps should be plotted instead of the default percentages

**Value**

A ggplot object using a barplot to show the distribution of the query regions across a given partition list.

**Examples**

```
p = calcPartitionsRef(vistaEnhancers, "hg19")
partPlot = plotPartitions(p)
partCounts = plotPartitions(p, numbers=TRUE)
```

---

plotQTHist	<i>Plot quantile-trimmed histogram</i>
------------	----------------------------------------

---

### Description

Given the results from calcWidth, plots a histogram with outliers trimmed.

### Usage

```
plotQTHist(
  x,
  EndBarColor = "gray57",
  MiddleBarColor = "gray27",
  quantile = NULL,
  bins = NULL,
  indep = FALSE,
  numbers = FALSE
)
```

### Arguments

x	Data values to plot
EndBarColor	Color for the quantile bars on both ends of the graph (optional)
MiddleBarColor	Color for the bars in the middle of the graph (optional)
quantile	Quantile of data to be contained in each end bar (optional) Quantiles must be under .2, optimal size is under .1
bins	The number of bins for the histogram to allocate data to. (optional)
indep	logical value which returns a list of plots that have had their bins calculated independently; the normal version will plot them on the same x and y axis.
numbers	a logical indicating whether the raw numbers should be displayed, rather than percentages (optional).

### Details

x-axis breaks for the frequency calculations are based on the "divisions" results from helper function calcDivisions.

### Value

A ggplot2 plot object

### Examples

```
plotQTHist(runif(500)*1000)
plotQTHist(list(q1=runif(500)*1000, q2=runif(500)*1000))
```

---

setB\_100*Example BED file read with rtracklayer::import*

---

**Description**

Example BED file read with rtracklayer::import

**Usage**

```
data(setB_100)
```

**Format**

GenomicRanges::GRanges

---

splitDataTable*Efficiently split a data.table by a column in the table*

---

**Description**

Efficiently split a data.table by a column in the table

**Usage**

```
splitDataTable(DT, split_factor)
```

**Arguments**

DT	Data.table to split
split_factor	Column to split, which can be a character vector or an integer.

**Value**

List of data.table objects, split by column

---

theme_blank_facet_label	<i>Clear ggplot face label.</i>
-------------------------	---------------------------------

---

**Description**

Usually ggplot2 facets are labeled with boxes surrounding the label. This function removes the box, so it's a simple label for each facet.

**Usage**

```
theme_blank_facet_label()
```

**Value**

A ggplot theme

---

TSS_hg19	<i>hg19 TSS locations</i>
----------	---------------------------

---

**Description**

A dataset containing chromosome sizes for Homo Sapiens hg38 genome assembly

**Usage**

```
data(TSS_hg19)
```

**Format**

A named vectors of lengths with one item per chromosome

**Source**

EnsDb.Hsapiens.v75 package

---

vistaEnhancers	<i>Example BED file read with rtracklayer::import</i>
----------------	-------------------------------------------------------

---

**Description**

Example BED file read with rtracklayer::import

**Usage**

```
data(vistaEnhancers)
```

**Format**

GenomicRanges::GRanges

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